



## CheKine™ Micro Urease Activity Assay Kit

Cat #: KTB3070

Size: 48 T/96 T

	<b>Micro Urease Activity Assay Kit</b>		
<b>REF</b>	<b>Cat #:</b> KTB3070	<b>LOT</b>	<b>Lot #:</b> Refer to product label
	<b>Applicable samples:</b> Animal and Plant Tissue, Cell, Bacteria, Serum, Plasma		
	<b>Storage:</b> Stored at 4°C for 12 months, protected from light		

### Assay Principle

Urease (UE) can hydrolyze urea to produce ammonia and carbonic acid. UE activity was positively correlated with organic matter content, total nitrogen and available nitrogen content, reflecting the nitrogen status. CheKine™ Micro Urease Activity Assay Kit provides a simple, convenient and rapid UE activity detection method, which is suitable for the detection of animal and plant tissue, cell, bacteria, serum, plasma and other samples. The detection principle is that the NH<sub>3</sub>-N produced by urease hydrolysis of urea is determined by indophenol blue colorimetry, and there is a characteristic absorption peak at 630 nm.

### Materials Supplied and Storage Conditions

Kit components	Size		Storage conditions
	48 T	96 T	
Extraction Solution	60 mL	120 mL	4°C
Reagent I	1	1	4°C
Reagent II	22 mL	44 mL	4°C
Reagent III A	0.4 mL	0.8 mL	4°C
Reagent III B	1.6 mL	3.2 mL	4°C
Reagent IV	2 mL	4 mL	4°C, protected from light
NH <sub>4</sub> Cl Standard (1 M)	0.2 mL	0.4 mL	4°C

### Materials Required but Not Supplied

- Microplate reader or visible spectrophotometer capable of measuring absorbance at 630 nm
- 96-well plate or microglass cuvette, precision pipettes, disposable pipette tips
- Ice maker, centrifuge, water bath
- Deionized water

- Homogenizer (for tissue samples)

## Reagent Preparation

**Extraction Solution:** Ready to use as supplied. Equilibrate to room temperature before use. Store at 4°C.

**Reagent I :** Before use, add 9 mL deionized water for 48T; add 18 mL deionized water for 96T; Fully dissolve it. Store at 4°C.

**Reagent II :** Ready to use as supplied. Equilibrate to room temperature before use. Store at 4°C.

**Reagent III:** Before use, pour liquid A into liquid B and mix it for use, and store the inexhaustible reagents at 4°C for a week.

**Reagent IV:** Ready to use as supplied. Equilibrate to room temperature before use. Store at 4°C, protected from light.

**Standard Curve Setting:** 1 M NH<sub>4</sub>Cl Standard was diluted to 1 mM with Extraction Solution. Dilute the 1 mM NH<sub>4</sub>Cl Standard with Extraction Solution to 500, 250, 125, 62.5, 31.25, 15.625 μM as indicated in the table below.

Num.	Volume of Standard	Volume of Extraction Solution (μL)	The Concentration of Standard (μM)
Std.1	200 μL of Standard	0	1,000
Std.2	100 μL of Std.1 (1,000 μM)	100	500
Std.3	100 μL of Std.2 (500 μM)	100	250
Std.4	100 μL of Std.3 (250 μM)	100	125
Std.5	100 μL of Std.4 (125 μM)	100	62.5
Std.6	100 μL of Std.5 (62.5 μM)	100	31.25
Std.7	100 μL of Std.6 (31.25 μM)	100	15.625

## Sample Preparation

**Note: Fresh samples are recommended. If the experiment is not carried out immediately, the samples can be stored at -80°C for 1 month.**

1. Tissues: Weigh 0.1 g tissue, add 1 mL Extraction Solution and homogenize on ice. Centrifuge at 8,000 g for 10 min at 4°C. Use supernatant for assay, and place it on ice to be tested.
2. Cell (bacteria): Collect 5×10<sup>6</sup> cell or bacteria into the centrifuge tube, wash with cold PBS, discard the supernatant after centrifugation; add 1 mL Extraction Solution to ultrasonically disrupt the bacteria 5 min (power 20% or 200 W, ultrasonic 3 s, interval 7 s, repeat 30 times). Centrifuge at 8,000 g for 10 min at 4°C. Use supernatant for assay, and place it on ice to be tested.
3. Serum (plasma): Direct detection.

**Note: It will be better to quantify the total protein with Protein Quantification Kit (BCA Assay), Cat #: KTD3001, if the content is calculated by protein concentration.**

## Assay Procedure

1. Preheat the microplate reader or ultraviolet spectrophotometer for more than 30 min, and adjust the wavelength to 630 nm, visible spectrophotometer was returned to zero with deionized water.
2. Enzymatic reaction (the following operations are performed in a 1.5 mL centrifuge tube).

Reagent	Test Tube (μL)	Control Tube (μL)
Sample	20	20
Reagent I	90	0
Deionized Water	0	90
Reagent II	190	190

Mix well, put it in a water bath at 37°C for 1 h, centrifuge at 10,000 g at 25°C for 10 min, and take the supernatant.

3. Dilute the supernatant 10 times (take 0.1 mL supernatant and add 0.9 mL deionized water).

4. Determination of ammonia (the following operations are performed in 96-well plates or microglass cuvette):

Reagent	Test Well (μL)	Control Well (μL)	Standard Well (μL)	Blank Well (μL)
Diluted Supernatant	80	80	0	0
Standard	0	0	80	0
Deionized Water	0	0	0	80
Reagent III	15	15	15	15
Reagent IV	15	15	15	15

Mix well and place 20 min at room temperature.

Deionized Water	90	90	90	90
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After rapid mixing, the absorbance value was determined by 630 nm, which was recorded as  $A_{\text{Test}}$ ,  $A_{\text{Control}}$ ,  $A_{\text{Standard}}$ ,  $A_{\text{Blank}}$ , calculated  $\Delta A_{\text{Test}} = A_{\text{Test}} - A_{\text{Control}}$ ,  $\Delta A_{\text{Standard}} = A_{\text{Standard}} - A_{\text{Blank}}$ .

**Note: Blank well and standard well only need to measure 1 time. In order to guarantee the accuracy of experimental results, need to do a pre-experiment with 2-3 samples. If  $\Delta A_{\text{Test}}$  is less than 0.001, increase the sample quantity appropriately. If  $\Delta A_{\text{Test}}$  is greater than 0.6, the sample can be appropriately diluted with Extraction Solution, the calculated result multiplied by the dilution factor, or decrease the sample quantity appropriately.**

## Data Analysis

**Note: We provide you with calculation formulae, including the derivation process and final formula. The two are exactly equal. It is suggested that the concise calculation formula in bold is final formula.**

1. Drawing of standard curve

With the concentration of the Standard Solution as the y-axis and the  $\Delta A_{\text{Standard}}$  as the x-axis, draw the standard curve. Substitute the  $\Delta A_{\text{Test}}$  into the equation to obtain the y value (μM).

2. Calculation of UE activity of samples

(1) Calculation of UE activity in serum (plasma):

Definition of unit: each mL serum (plasma) produces 1 μmol of NH<sub>3</sub>-N per min is defined as an enzyme activity unit.

$$\text{UE (U/mL)} = y \times (V_{\text{Total reaction}} \div V_{\text{Sample}}) \times 10 \div T \div 1,000 = \mathbf{0.0025y}$$

(2) Calculation of UE activity in tissues

a. Calculated according to the concentration of sample protein

Definition of unit: 1 μmol of NH<sub>3</sub>-N produced per mg tissue protein per min is defined as an enzyme activity unit.

$$\text{UE (U/mg prot)} = y \times (V_{\text{Total reaction}} \div V_{\text{Sample}}) \times 10 \div T \div 1,000 \div \text{Cpr} = \mathbf{0.0025y \div \text{Cpr}}$$

b. Calculated according to sample fresh weight

Definition of unit: 1 μmol of NH<sub>3</sub>-N produced per g of tissue per min is defined as an enzyme activity unit.

$$\text{UE (U/g fresh weight)} = y \times (V_{\text{Total reaction}} \div V_{\text{Sample}}) \times 10 \times V_{\text{Total sample}} \div T \div 1,000 \div W = \mathbf{0.0025y \div W}$$

c. Calculated according to cell (bacterial) density

Definition of unit: every 1 million cells (bacteria) produces 1 μmol of NH<sub>3</sub>-N per min is defined as an enzyme activity unit.

$$\text{UE (U/10}^6) = y \times (V_{\text{Total reaction}} \div V_{\text{Sample}}) \times 10 \times V_{\text{Total sample}} \div T \div 1,000 \div n = \mathbf{0.0025y \div n}$$

Where:  $V_{\text{Total reaction}}$ : Total volume of reaction system, 0.3 mL;  $V_{\text{Sample}}$ : Add sample volume, 0.02 mL; T: Reaction time, 60 min; 1,000: Unit conversion coefficient, 1L=1,000 mL; Cpr: Protein concentration, mg/mL;  $V_{\text{Total sample}}$ : Extract solution volume, 1 mL; W: Sample quality, g; n: Total cell (bacteria), million.

## Typical Data

Typical standard curve:

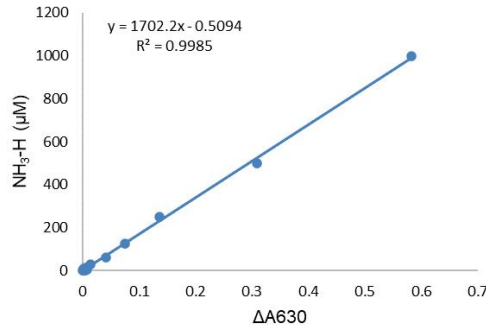


Figure1. Standard Curve for NH<sub>3</sub>-N.

Examples:

1. Take 0.1 g mouse liver and use 96-well plate to calculate  $\Delta A_{\text{Test}} = 0.206 - 0.09 = 0.116$ ,  $y = 196.985$ . The content calculated according to the sample quality is as follows:

UE (U/g fresh weight) =  $0.0025 \times 196.985 \div 0.1 = 4.925$  U/g.

2. Take 5 million Jurkat cells and use 96-well plate to calculate  $\Delta A_{\text{Test}} = 0.110 - 0.044 = 0.066$ ,  $y = 111.753$ . The content calculated according to the cell density is as follows:

UE (U/10<sup>6</sup>) =  $0.0025 \times 111.753 \div 5 = 0.056$  U/10<sup>6</sup>.

3. Take 20 μL bovine serum and use 96-well plate to calculate  $\Delta A_{\text{Test}} = 0.210 - 0.093 = 0.117$ ,  $y = 198.689$ . The content calculated according to the liquid volume is as follows:

UE (U/mL) =  $0.0025 \times 198.689 \div 5 = 0.497$  U/mL.

## Recommended Products

Catalog No.	Product Name
KTB3040	CheKine™ Micro Glutamate Synthase (GOGAT) Assay Kit
KTB3041	CheKine™ Micro Glutamic Acid Dehydrogenase (GDH) Assay Kit
KTB3050	CheKine™ Micro Water and Soil Nitrite Content Assay Kit
KTB3051	CheKine™ Micro Food Nitrite Assay Kit

## Disclaimer

The reagent is only used in the field of scientific research, not suitable for clinical diagnosis or other purposes.